

# Development and Validation of Stability-Indicating HPLC Method for Simultaneous Determination of Emtricitabine and Tenofovir Disoproxil Fumarate in Bulk and their Tablet Dosage Form

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## Abstract:

A reversed-phase high performance liquid chromatographic method is simple, precise, reliable, rapid and reproducible method, developed and validated for the simultaneous estimation of Emtricitabine and Tenofovir disoproxil fumarate. Chromatography is carried out isocratically on Intersil C18 (33mm X 4.6 mm) column with a mobile phase composed of acetonitrile: phosphate buffer (80:20 v/v) at a flow rate of 2.0ml/min. Detected with the help of UV detector at wavelength 260 nm. Linearity, precision, accuracy, specificity and ruggedness are the parameters analysed as per the ICH Q2(R1) guidelines. The retention times of Emtricitabine and Tenofovir are 2.1 min and 14.3 min respectively. The 1% recoveries of Emtricitabine and Tenofovir are found to be 99.5%, 99.8% respectively. The method developed was found to be accurate, precise, selective and rapid for simultaneous estimation of Emtricitabine and Tenofovir in pharmaceutical dosage forms.

**Key words**—Reverse phase HPLC, EMTRICITABINE, TENOFOVIR DISOPROXIL FUMERATE, ISOCRATIC, INTERSIL C18, acetonitrile, phosphate buffer.

## I. INTRODUCTION:

**Emtricitabine:** Chemically 4-amino-5-fluoro-1-[(2R,5S)-2-(hydroxymethyl)-1,3-oxathiolan-5-yl]-1,2-dihydropyrimidin-2-one [1]. And with molecular formula of C<sub>8</sub>H<sub>10</sub>N<sub>3</sub>O<sub>3</sub>S and a molecular weight- 247.24 g/mol. Emtricitabine is a synthetic nucleotide analogue of cytidine. It has solubility in 112mg/ml of distilled water at 25 °C and it is freely soluble in various aqueous solutions, slightly soluble in acetonitrile and very slightly soluble in isopropyl acetate, soluble in methanol. Emtricitabine is used in the treatment of HIV. **Emtricitabine** is an analogue of cytidine. The mechanism of action of the drug was by inhibiting reverse transcriptase, the enzyme that copies HIV RNA into new viral DNA. Side effects like diarrhea, headache, trouble sleeping, Darkening skin colour on palms of hand and soles of feet may occur.

**Tenofovir Disoproxil:** Chemically it is 9-[(R)-2-[[[isopropoxycarbonyl]-oxy] methoxy] phosphinyl] methoxy] propyl] adenine fumarate [4e7]. It has molecular formula C<sub>9</sub>H<sub>14</sub>N<sub>5</sub>O<sub>4</sub>P and a molecular weight 635.52g/mol. It has Solubility in 13.4mg/ml of distilled water at 25°C, it is freely soluble in dimethylformamide, soluble in methanol, acid hydrochloride (0.1N HCl), ethanol, sparingly soluble in acetone, isopropanol, water,

slightly soluble in acetonitrile, ethylacetate, insoluble in dichloromethane, hexane. It is also used for the treatment of HIV. **Tenofovir disoproxil Fumarate** :Tenofovir disoproxil Fumarate exhibits activity against HIV-reverse transcriptase.

Side effects include Dizziness, diarrhoea, headache, trouble sleeping.

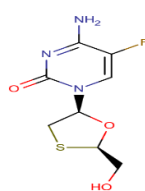


Fig no:1 Emtricitabine

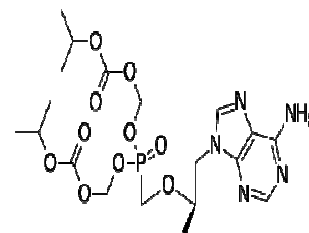


Fig no:2 Tenofovir Disoproxil

The literature survey reveals that there was only one HPLC [3] method was developed in combination of Emtricitabine and Tenofovir disoproxil in bulk and dosage forms. There were no reported analytical methods for simultaneous estimation of Emtricitabine and Tenofovir disoproxil in bulk and their combined dosage forms in presence of their

degradation products. Hence it is an attempt to develop stability indicating specific, sensitive, accurate and precise RP-HPLC method for simultaneous estimation of these drugs. This method was validated as per the guidelines of ICH Q2.

## II. MATERIALS AND METHODS:

### Instruments used:

s.no	Name of instrument	Make	Model	Software
1	HPLC	AGILENT	UV Detector	Empower3
2	Weighing Balance	SARTORIUS	BSA224S-CW	N/A
3	p <sup>H</sup> meter	LABINDIA	AD102U	N/A
4	Sonicator, Centrifuge	ENERTECH	SE60US	N/A

Table no :1 Instruments used in the method

### Chemicals used:

S.No.	Chemicals/Reagents	Make/Grade
1	Methanol	Merck (HPLC-Grade)
2	Acetonitrile	Merck (HPLC-Grade)
4	Potassium dihydrogen Orthophosphate	Merck (GR-Grade)
5	O-Phosphoric acid	Merck (GR-Grade)

Table no:2 Chemicals used in the method

### Working reference standards used:

S.No	Name	Batch number	%Potency (As is basis)
1	Emtricitabine	VJR(681)187	99.2%
2	Tenofovir Disoproxil fumarate	KSC(708)164	98.9%
3	placebo	AS(895)166P	99.1%
4	Tablets	AS(895)	98.6%

Table no:3 Standards used for the reference

### Selection of Buffer and Mobile phase:

Initially development was started with a buffer which was used for Emtricitabine, Tenofovir Disoproxil Fumarate method (0.01M KH<sub>2</sub>PO<sub>4</sub>+1g-1-Octane Sulfonic Acid pH3.0). But to separate impurities from analytes buffer 0.01M KH<sub>2</sub>PO<sub>4</sub>+1g-1-Octane Sulfonic Acid pH3.0 with Acetonitrile and Methanol with different gradient programmers were tried. Out of this Buffer pH 3.0 with Acetonitrile: Methanol (80:20) was finalized Based on peak parameters like resolution power, retention time, good peak shape, peak tailing and no blank interference.

### Selection of HPLC column:

Various HPLC columns polar packing material like Hypersil BDS C18 (100 x 4.6mm, 5 μm), Hypersil BDS C18 (250 x 4.6mm, 5 μm), μ Bond pack column (300x4.6mm, 10 μm) used for this study. Out of these Inertsil C8 33\*4.6mm, 3μm was finalized based on peak parameters like retention time, peak shape, resolution power and peak tailing.

## OPTIMIZED HPLC METHOD

### Preparation of Buffer:

Transfer about 2.72g of Potassium dihydrogen Phosphate into a beaker containing 1000 mL of Milli-Q water Adjust pH of the solution to 4.0±0.05 with diluted Ortho phosphoric acid solution. Filter the solution through 0.45μm membrane filter.

Transfer about 3.4g of Potassium dihydrogen Phosphate into a beaker containing 1000 mL of Milli-Q water Adjust pH of the solution to 3.0±0.05 with diluted Ortho phosphoric acid solution. Filter the solution through 0.45μm membrane filter

### Preparation of Mobile phase-A: Use Buffer (pH 4.0)

**Preparation of Mobile Phase-B:** Prepare a degassed mixture of buffer (pH 4.0) and Acetonitrile in the ratio of 20:80 (%v/v)

**Preparation of diluent:** Prepare a degassed mixture of buffer (pH 3.0) and Methanol in the ratio of 50:50 (%v/v)

### Preparation of Emtricitabine and Tenofovir disoproxil fumarate standard stock solution:

Accurately weigh and transfer about 40mg of Emtricitabine and 60mg Tenofovir disoproxil fumarate working standard into a 50mL volumetric flask. Add about 30mL of Methanol and sonicate to dissolve. Dilute to volume with Methanol and mix.

Transfer 10mL of above solution into a 50mL volumetric flask, dilute to volume with diluent and mix. Filter the solution through 0.45µm membrane filter.

#### Preparation of sample solution:

##### For 200/300mg Tablets

Transfer 5tablets in to a 500mL volumetric flask, add about 200mL of buffer (pH 3.0) and mechanical shaker for 1hour 150rpm. Add Methanol up to 1cm bellow the mark and sonicate for not less than 30minutes with occasional shaking {maintain the sonicator bath temperature between 20 to 25°C). Dilute to volume with Methanol and mix. Centrifuge a portion of the solution at 5000rpm for about 5minutes.

Transfer 4mL of above solution into a 50mL volumetric flask, dilute to volume with diluent and mix. Filter the solution through 0.45µm membrane filter and discard first few ml of the filtrate.

#### Procedure

Equilibrate the column for not less than 60 minutes with initial mobile phase composition at a flow rate of 1.0mL/minute.

Separately inject 10uL of Blank (diluent), Standard solution (five injections) and Sample solution into the chromatographic system. Record the chromatograms and measure the peak responses.

#### VALIDATION OF OPTIMIZED METHOD:

The proposed method for the assay of Emtricitabine, Tenofovir disoproxil fumarate and tablet dosage form was subjected to validation to check its suitability for routine analysis.

##### A. SPECIFICITY:

##### Blank interference

Diluent was prepared as per the test method and injected into the chromatographic system and the chromatograms were recorded.

##### Placebo Interference:

Diluent was injected into Chromatographic system and found no interference. Placebo sample was prepared by taking the placebo equivalent to about the weight portion of test preparation and injected into the HPLC system.

##### Impurity Interference:

All the related compounds are injected into the HPLC system and recorded the chromatograms.

##### B. LINEARITY:

The linearity of the method was demonstrated over the concentration range of 50% - 150% of the target concentration. Aliquots of 50%, 70%,80%,90% 100%,110% ,130% and 150% were prepared from standard stock solution.

##### Procedure

Standard solutions of 50 – 150% concentration were injected separately into the chromatographic system and the chromatograms were recorded. Peak areas were recorded for each injected concentration of drugs and the calibration curves, concentration vs. peak area were constructed for the drugs.

##### C. PRECISION:

The precision of the method was determined by system precision and method precision using 100% standard and sample solutions.

##### System precision

The system precision was established by injecting six replicate injections of standard solution in to the chromatographic system and the chromatograms were recorded.

##### Method precision

Six assay samples of drug product at 100% of the sample concentration were prepared and injected into the chromatographic system and the chromatograms were recorded.

The system and method precision were calculated by using formulae:

$$\text{Standard deviation, S.D} = \sqrt{\frac{\sum_{i=1}^{i=n} (x_i - \bar{x})^2}{N - 1}}$$

$$\% \text{ RSD} = \frac{\text{S.D}}{\bar{x}} \times 100$$

Where,  $\bar{X}$  = Mean

N = No. of samples

S. D=Standard deviation

%RSD = Percent relative S.D

#### D. ACCURACY:

The solutions were prepared in replicate at levels 50%,100% and 150% of test concentration using Emtricitabine and Tenofovir disoproxil fumarate drug substance and Emtricitabine and Tenofovir disoproxil fumarate 200/300 tablets placebos per the test method and injected each solution into HPLC as per methodology.

**Acceptance criteria:** Recovery should be between 98.0% to 102.0%

#### E. ROBUSTNESS:

As part of evaluation of robustness, deliberate changes were made in the flow rate, Organic phase modifications and wavelength to evaluate the impact on the method.

#### Procedure

##### Effect of variation of flow rate

Standard solution prepared as per the test method was injected into the chromatographic system maintaining flow rates, less flow(1.8mL/min), more flow(2.2mL/min) and actual flow (2.0mL/min).

##### Effect of variation of Organic phase

Standard solution prepared as per the test method was injected into the chromatographic system maintaining flow rates, 2% less organic phase ,2% more organic phase and actual organic phase.

##### Effect of variation of wavelength

Standard solution prepared as per the test method was injected into the chromatographic system maintaining flow rates, less wavelength (255nm), more wavelength(265nm) and actual wavelength(260nm).

#### Stress degradation studies

Stress degradation study was conducted in acid, base, peroxide and homogeneity of the peak was assessed in terms of peak purity.

#### Filter validation

Filter validation was performed by filtering standard and sample solutions with different filters.

#### Solution stability

Solution stability was performed by injecting standard and sample solution after 12Hrs,24Hrs at 6° c.

### III. RESULTS AND DISCUSSION:

#### Optimised Chromatographic Conditions:

A Gradient Rapid and simple RP-HPLC method was developed and validated for the simultaneous estimation of drugs. Injection volume is set to 10µl. Mobile phase consisting of degassed mixture of phosphate dihydrogen buffer (pH 4.0) and Acetonitrile in the ratio of 20:80 (%v/v) was set with gradient programming for 25 minutes. chromatographic conditions were optimised for mobile phase using inertsilC8(33mm×4.6mm,3µm) column at a flow rate of 2.0ml/min. effluents was detected at 260nm by variable wavelength UV detector. column compartment temperature was 30°C.

#### Validation parameters:

##### a) Specificity:

The chromatograms of **Emtricitabine, Tenofovir DF** were taken and shown in Figure-3, The peak areas were observed.

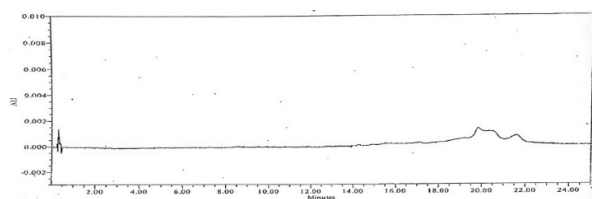


Fig no :3 HPLC Chromatogram of Placebo

**Observation:** No interference was observed

**Acceptance:** No peak should not be found at the retention time of main peak

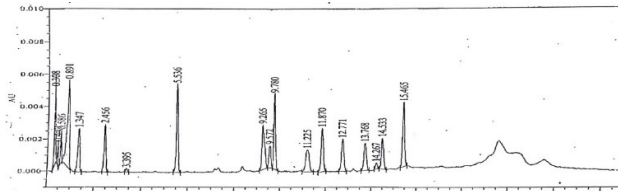


Fig no:4 Impurity Interference Chromatogram of Emtricitabine, Tenofovir Disoproxil Fumarate

Observation :No interference was observed

Acceptance: No peak should be found at the retention time of main peak

**b) Linearity**

The chromatograms of Emtricitabine, Tenofovir Disoproxil Fumarate were taken and shown in Figures 7.16, 7.17, 7.18, 7.19, 7.20, 7.21, 7.22, 7.23. The peak areas were observed. The correlation coefficient@ 0.9998 and %y intercept - 0.3for emtricitabine, and for tenofovir disoproxil fumarate was 0.9998 and %y intercept -0.5

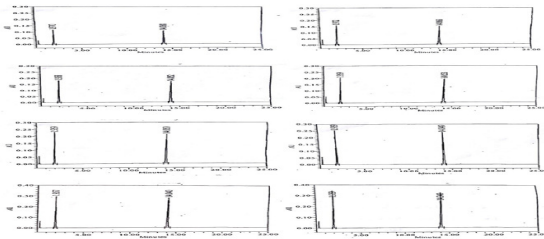
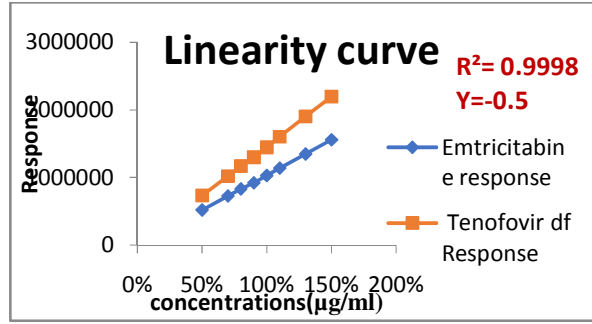


Fig no: 5 HPLC chromatographs for linearity

50%,70%,80%,90%,100%,110%,130%,150% respectively

Table no: 5 Linearity data of Emtricitabine and Tenofovir disoproxil

S.NO	Linearity level	Emtricitabine response	Tenofovir df Response
1	50%	521048	734215
2	70%	727336	1021828
3	80%	831900	1170230
4	90%	922284	1299237
5	100%	1028697	1446794
6	110%	1139566	1604068
7	130%	1348370	1902641
8	150%	1558431	2197338
	Correlation coefficient	0.9998	0.9998
	%y-intercept	-0.3	-0.5



Graph no: 1 Linearity curve of Emtricitabine and Tenofovir disoproxil responses

**c) Precision**

Six sample solutions were prepared. The chromatograms were observed and shown in Figures

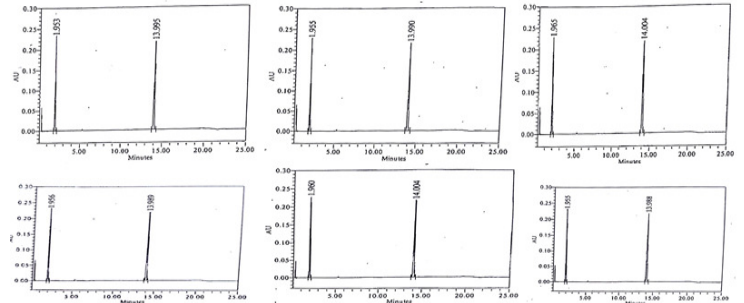


Fig no: 6 HPLC chromatogram of precision

S.NO	Sample Name	Emtricitabine	Tenofovir DF
1	Precision sample-1	1029888	1447111
2	Precision sample-2	1027490	1447505
3	Precision sample-3	1033097	1452702
4	Precision sample-4	1034906	1455942
5	Precision sample-5	1036324	1457164
6	Precision sample-6	1037325	1453083
	Mean	1033172	1453083
	Std. Dev.	3828	4830
	%RSD	0.4	0.3

Table no: 6 System precision data of Emtricitabine, Tenofovir df

S.NO	Sample Name	Emtricitabine	Tenofovir DF
1	Precision sample-1	101.1	101.3
2	Precision sample-2	99.85	102.6
3	Precision sample-3	101.3	101.7
4	Precision sample-4	100	100.6

5	Precision sample-5	101.2	101.5
6	Precision sample-6	100.4	100.8
Mean		100.6	101
%RSD		0.6	0.5

Table no :6 Method precision data of Emtricitabine and Tenofovir df

**Observation:** The % RSD and % Assay of Emtricitabine, Tenofovir DF are found to be 0.6, 0.5, and 100.6, 101 respectively

**Acceptance criteria:** The % RSD should be NMT 2.0% and Average % Assay should be between 95-105% of labelled amount.

**d) Accuracy**

To determine the accuracy of the method recovery was performed by standard addition method. To pre-analysed sampled, known amount of standards. Emtricitabine, Tenofovir DF were spiked in different concentrations. The recoveries of Emtricitabine, Tenofovir DF at three levels 50%, 100% and 150%.

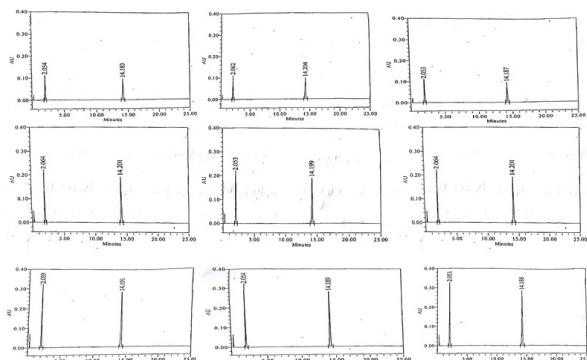


Fig no 7 Accuracy data of Emtricitabine and Tenofovir df

S.NO	Accuracy level	% Recovery	Mean recovery	Overall mean recovery
1	Emtricitabine	99.8, 99.2, 100.5	99.8	99.5
	100%	99.4, 99.4, 99.1	99.3	
	50%	99.4, 98.8, 99.8	99.3	
2	Tenofovir DF-50%	100.1, 99.4, 101.1	100.2	

5	100%	99.6, 99.4	99.6, 99.5	99.8
6	50%	99.8, 100.2	99.3, 99.8	

Table no: 7 Accuracy Data

**Observation:** Accuracy was found to be 99.5% and 99.8

**e) Robustness:**

**Robustness conditions**

To prove the Robustness of the method, the following parameters were also verified.

Flow,pH,Wavelength, % of organic in mobile phase

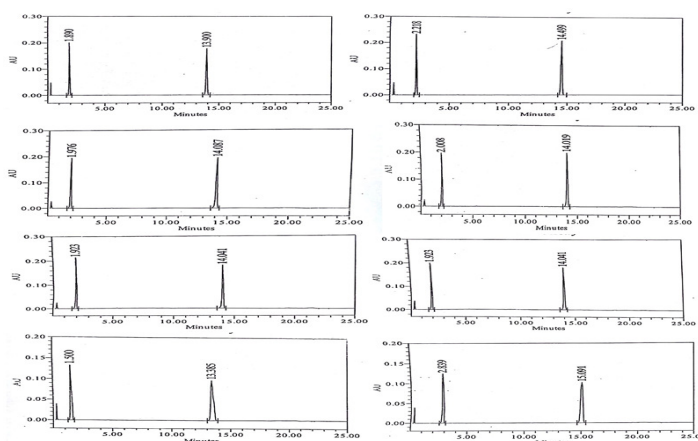


Fig no: 8 Chromatograms of Flow rate (+), Flow rate (-), PH (+), PH (-), Wave length(+) and Wave length(-) respectively.

Parameter	Variation	% RSD	
		Emtricitabine	Tenofovir DF
STP	-	0.5	0.5
Flow Rate	-10%	0.4	0.4
	+10%	0.3	0.3
Wavelength	-5nm	0.5	0.5
	+5nm	0.5	0.5
% Organic in mobile phase	-2% absolute	0.2	0.2
	+2% absolute	0.4	0.4

pH	-0.2 units	0.6	0.5	Solution stability was performed by injecting standard and sample solution after 3 Hrs at room temperature 25°C.
	+0.2 units	0.2	0.2	

Table no: 8 Robustness data of Emtricitabine and Tenofovir DF

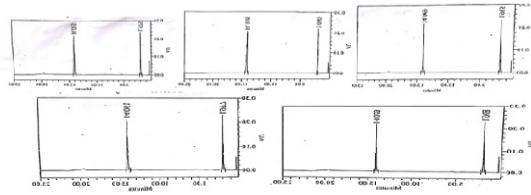


Fig no:10 Solution stability chromatographs at 25°C

i. DEGRADATION STUDY:

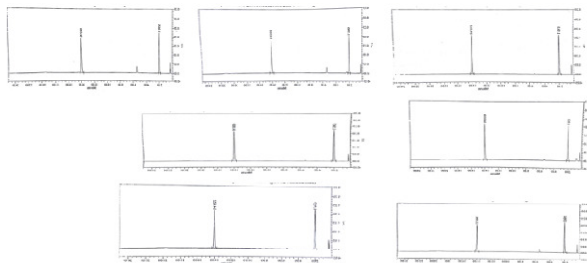


Fig no :9 Chromatograms for Un degradation, Acid degradation, Basedegradation, Peroxidedegradation, Photolyticdegradation, Thermaldegradation, Humidity degradation.

Solution stability was performed by injecting standard and sample solution after 12Hrs at refrigerator temperature at 6°C.

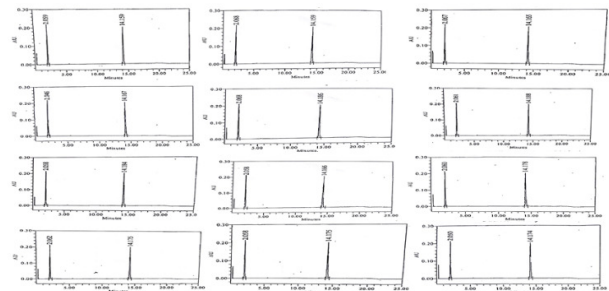


Fig no:11 Solution stability chromatographs at 6°C.

Sample Name	Condition	Purity angle	Purity Threshold	Condition	% Difference	
					Emtricitabine	Tenofovir DF
Emtricitabine	Acid Degradation	0.032	0.252	Room temperature up to 3 Hours (~25°C) Refrigerator condition up to 12 Hours (~6°C)	0.5	0.1
	Base Degradation	0.036	0.243			
	Peroxide Degradation	0.036	0.245		0.7	0.3
	Photolytic Degradation	0.038	0.250			
	Thermal Degradation	0.035	0.253			
	Humidity Degradation	0.037	0.244			
Tenofovir DF	Acid Degradation	0.045	0.260	Refrigerator condition up to 12 Hours (~6°C)	0.7	0.3
	Base Degradation	0.038	0.244			
	Peroxide Degradation	0.034	0.242		0.7	0.3
	Photolytic Degradation	0.031	0.244			
	Thermal Degradation	0.056	0.262			
	Humidity Degradation	0.039	0.242			

Table no: 10 Solution stability data of Emtricitabine, Tenofovir DF

Observation: %Difference to be found 0.5,0.1and 0.7,0.3 for Emtricitabine, Tenofovir DF in Sample Respectively.

Acceptance criteria: %RSD of Assay difference to the initial value should not be more than 2.0%

Table no: 9 Data of forced degradation

Observation: Purity angle is less than purity threshold in all the stress conditions.

Acceptance Criteria: Purity angle should be less than purity threshold.

ii. SOLUTION STABILITY

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16. MN- Trinath, BM Gurupadayya\* Shiva Prasad, Shilpa Kache
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